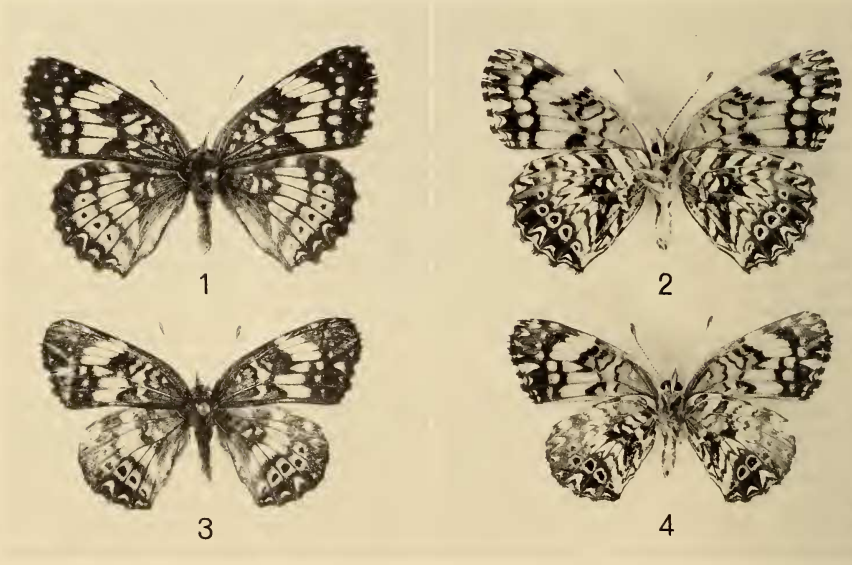
retained by W. H. Edwards and now is with his collection in the Carnegie Museum of Natural History, Pittsburgh, Pa. Its locality of capture at this time can be given no more closely than vicinity of Independence, Inyo Co., California, from a colony now extinct.

F. MARTIN BROWN, *Wright-Ingraham Institute, Colorado Springs, Colorado 80911.*

Journal of the Lepidopterists' Society
37(1), 1983, 80–81

A NEW FOOD PLANT RECORD FOR *CHLOSYNE GORGONE CARLOTA* (REAKIRT) (NYMPHALIDAE)

Host plants recorded for the larval stages of *Chlosyne gorgone carlota* (Reakirt) include a wide variety of genera in several families. While most authors (Klots, 1951, Field Guide to the Butterflies, Houghton Mifflin Co.; Forbes, 1960, Lepidoptera of New York and Neighboring States, Part IV, Cornell Univ. Agr. Expt. Sta. Memoir 371; Ehrlich & Ehrlich, 1961, How to Know the Butterflies, Wm. C. Brown Co.; Tietz, 1972, An Index to the Described Life Histories, Early Stages, and Hosts of the Macrolepidoptera of the Continental United States and Canada, Allyn Mus. Entomol., Sarasota, FL; Johnson, 1972 (1973), J. Res. Lepid. 11(1):1–64) list *Aster* spp. and *Helianthus* spp. (Compositae) as the primary food plants, *C. g. carlota* larvae have also been re-



FIGS. 1–4. Female specimens of *Chlosyne gorgone carlota* from Missouri: **1, 2,** female showing typical markings of this species, dorsal and ventral views, respectively; **3, 4,** female reared from *Ambrosia trifida*, dorsal and ventral views, respectively.

corded from *Lippia lanceolata* and *L. nodiflora* (Verbenaceae) (Kimball, 1965, Arthropods of Florida and Neighboring Land Areas, Vol. 1, Lepidoptera of Florida, Florida Dept. Ag.), *Lysimachia* sp. (Primulaceae) (Harris, 1972, Butterflies of Georgia, Univ. of Oklahoma Press), and *Eriogonum* sp. (Polygonaceae) (Tietz, ibid.).

In Missouri, *C. g. carlota* larvae have been recorded from *Helianthus annuus* L. (Compositae) (J. R. Heitzman, pers. comm.) and *Linaria vulgaria* Hill (Scrophulariaceae) (Masters, 1969, J. Kansas Entomol. Soc. 42(2):133–144). Here I report giant ragweed, *Ambrosia trifida* L. (Compositae), as a new food plant for *C. g. carlota*. *A. trifida* L. is an annual, monoecious weed common to fertile moist soils, bottom lands, alluvium, and waste places.

On 9 August 1977, while collecting along a roadside bank of State Rt. 10 (3.2 km west of Richmond, MO), I found thirteen larvae that were unfamiliar to me on the leaves of three separate giant ragweed plants. Due to the uniformity in larval size and the close proximity of the plants to each other (within 0.5 m of each other), the infestation was probably the result of a single oviposition. The thirteen larvae, along with an ample supply of the food plant, were collected and taken to my home to be reared. Six larvae were preserved on 9 August 1977; the remaining seven larvae were allowed to feed. On 10 August 1977 one larva pupated, emerging 11 days later. The single adult, identified as a female *C. g. carlota* by J. R. Heitzman, is slightly smaller and darker than the typical female of the species (Figs. 1–4), which may have resulted from the rearing. The six remaining larvae pupated on 11 August 1977 and were preserved.

I am especially grateful to J. Richard Heitzman for identifying the specimen and for reviewing this manuscript. Thanks are also extended to Dr. Charles V. Covell, Jr., Dr. Stephen Clement and to Ms. Candy Fogg for their critical reviews and to Mr. Glenn Berkey of the Ohio Agricultural Research and Development Center, Wooster, for the photographs.

DAVID C. IFTNER, *2161 Heatherfield Avenue, Worthington, Ohio 43085.*

Journal of the Lepidopterists' Society
37(1), 1983, 81–82

A NATURAL OCCURRENCE OF INTER-TRIBAL COPULATION IN THE PAPILIONIDAE

Among the Lepidoptera numerous prezygotic isolating mechanisms (Mayr, 1970, Population, Species and Evolution, Belknap Press, Cambridge, MA) operate to prevent interspecific mating. At times, however, these isolating mechanisms appear to break down. While breakdowns occur rarely between closely related species, breakdowns between distantly related and phenotypically distinct species are quite exceptional. A salient example of such a breakdown was observed on 15 April 1980 in the Kirby State Forest near Kountze, Tyler County, Texas.

Swallowtail diversity in Kirby Forest and throughout the upland savannah of southeast Texas is high; at least six different species are resident (Rausher, in preparation). *Eurytides marcellus* (Papilionidae: Graphiini) and *Battus philenor* (Papilionidae: Troidini) fly sympatrically in the open pine uplands. The two species are temporally synchronous, adults flying commonly between mid-March and mid-April. During this period females spend much of their time searching among the herbaceous vegetation for larval food plants. For *B. philenor*, these are *Aristolochia reticulata* and *A. serpentaria*, small erect perennial herbs in the family Aristolochiaceae (Rausher, 1978, Science 200: 1071–1073). The larval food plant of *E. marcellus*, in contrast, is *Asimina parviflora*, an annonaceous shrub that grows in east Texas to a maximum height of 2–3 ft. Males of each species fly through the pine uplands, approach females while they are ovipos-